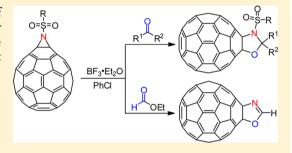


# BF<sub>3</sub>·Et<sub>2</sub>O-Catalyzed Formal [3 + 2] Reaction of Aziridinofullerenes with Carbonyl Compounds

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Supporting Information

**ABSTRACT:** The BF $_3$ ·Et $_2$ O-catalyzed formal [3+2] reaction of aziridinofullerenes with various carbonyl compounds for the easy preparation of fullerooxazolidines has been developed. Moreover, the reaction of aziridinofullerene with ethyl formate affords the simplest fullerooxazole without substituent.



fullerene derivatives, which can be easily synthesized from

azides, 4 chloramines, 5 sulfilimines, 6 iminophenyliodinanes, 7 and

N,N-dihalosulfonamides. We and Gan also reported a method

to prepare the aziridinofullerenes from amines promoted by

hypervalent iodine reagents.9 Although azafulleroid (isomer of

aziridinofullerene) is widely used in the synthesis of open-cage fullerene or azafullerene  $(C_{59}N)$ , <sup>10</sup> the transformation of

aziridinofullerene has seldom been investigated. The Minakata

and Itami groups used aziridinofullerene as a versatile platform

for the preparation of functionalized fullerenes by performing

the reactions of it with aromatic compounds or bifunctional

nucleophiles under acid conditions. 5b,7 In this transformation,

the generation of product is accompanied with the loss of

sulfonamide. In order to reserve the  $N_1$ -unit in the product, the

[3 + 2] reaction of masked 1,3-dipoles generated from Lewis

acid-catalyzed C-N bond cleavage with dipolarophiles is a

possible way. However, it is known to all that the C<sub>60</sub> moiety as

an electron-deficient alkene is not beneficial to the formation of

carbon cation, 11 which means that such a [3 + 2] reaction of

aziridinofullerene remains a challenge (Scheme 1). Most

recently, the Minakata group reported the Lewis base but not the usually used Lewis acid catalyzed ring expansion of

aziridinofullerene with  $CO_2$  and isocyanates.<sup>12</sup> In continuation of our interest in fullerene chemistry,<sup>13</sup> we reported here the  $BF_3 \cdot Et_2O$ -catalyzed the formal [3 + 2] reaction of aziridinofullerenes with carbonyl compounds for the easy preparation of

hemical modification of fullerene is still an attractive research field for designing more fullerene derivatives with unique physical, chemical, and biological properties. Among the numerous developed methods for preparation of functionalized fullerene, most of the derivatives are prepared directly from C<sub>60</sub>. However, some derivatives cannot be easily synthesized through a one-step reaction. Thus, the investigation of the further transformation starting from fullerene derivatives remains under development. Aziridine as a highly strained three-membered cyclic amine is one of the most important precursors to generate 1,3-dipoles or masked 1,3-dipoles through C-C or C-N bond cleavage, which can be trapped by various dipolarophiles for the preparation of nitrogencontaining five-membered-ring heterocycles.<sup>2</sup> The predominant bond cleavage depends mainly on the electronic properties of the substituent on the C-atom of aziridine ring.<sup>2,3</sup> The electronwithdrawing group is beneficial to the C-C bond cleavage, and the electron-donating group prefers the C-N bond breaking due to the stabilization of carbanion and carbon cation by delocalization, respectively (Scheme 1). Aziridinofullerene is one of the most important classes of nitrogen-containing

## Scheme 1

stablization of the anion or cation with R group fullerene cation, not stable

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oxazolidine-fused fullerene derivatives.

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We started our study by choosing N-tosylaziridinofullerene  ${\bf 1a}$  and benzaldehyde  ${\bf 2a}$  as the model substrates to test the possibility of formal [3+2] cycloaddition reaction. In order to avoid the reaction between  ${\bf 1a}$  and toluene,  ${}^{5b,14}$  the chlorobenzene was chosen as the solvent. Various commonly used Lewis acids including  $Sc(OTf)_3$ ,  $Cu(OTf)_2$ ,  $Zn(OTf)_2$ ,  $Sn(OTf)_2$ ,  $Cu(ClO_4)_2$ ,  $Cu(acac)_2$ ,  $Ni(acac)_2$ ,  $AgClO_4$ , and  $BF_3$ ·  $Et_2O$  were evaluated. Although  $Sc(OTf)_3$ ,  $Cu(OTf)_2$ , and  $Zn(OTf)_2$  have been reported to effectively catalyze the reaction of 2-aryl-N-tosylaziridine with aldehydes or ketones, in our hand all of them did not show any catalytic activity.  $Sn(OTf)_2$ ,  $Cu(ClO_4)_2$ ,  $Cu(acac)_2$ ,  $Ni(acac)_2$ , and  $AgClO_4$  could not catalyze the reaction either.  $BF_3$ · $Et_2O$  was the sole efficacious catalyst for this transformation to give the desired  $C_{60}$ -fused oxazolidine product  ${\bf 3aa}$  (Table 1, entry 9).

Table 1. Screening of the Reaction Conditions<sup>a</sup>

| citity | Dewis acid            | 14/24/1/11 | tillic | yicia (70) |
|--------|-----------------------|------------|--------|------------|
| 1      | $Cu(OTf)_2$           | 1:1.5:1    | 24 h   | 0          |
| 2      | $Zn(OTf)_2$           | 1:1.5:1    | 24 h   | 0          |
| 3      | $Sn(OTf)_2$           | 1:1.5:1    | 24 h   | 0          |
| 4      | $Sc(OTf)_3$           | 1:1.5:1    | 24 h   | 0          |
| 5      | AgClO <sub>4</sub>    | 1:1.5:1    | 24 h   | 0          |
| 6      | $Cu(ClO_4)_2$         | 1:1.5:1    | 24 h   | 0          |
| 7      | $Mg(ClO_4)_2$         | 1:1.5:1    | 24 h   | 0          |
| 8      | Cu(acac) <sub>2</sub> | 1:1.5:1    | 24 h   | 0          |
| 9      | $BF_3 \cdot Et_2O$    | 1:1.5:2    | 90 min | 67         |
| 10     | $BF_3 \cdot Et_2O$    | 1:1.5:5    | 15 min | 88         |
| arr 1  |                       |            | . 1    | 0.00 1.0   |

<sup>a</sup>Unless notification, the reactions were carried out with 0.02 mmol of 1a and proper additives in 2.5 mL of dry chlorobenzene at room temperature. <sup>b</sup>Isolated yield.

Increasing the amount of  $BF_3 \cdot Et_2O$  from 1.5 to 5 equiv had a noticeable acceleration effect on the reaction and led to 88% yield of 3aa within 15 min (Table 1, entry 10).

With the optimal reaction conditions in hand, we next studied the scope of the BF<sub>3</sub>·Et<sub>2</sub>O-catalyzed formal [3 + 2] cycloaddition reaction of N-tosylaziridinofullerene 1a with various carbonyl compounds (Table 2). All the carbonyl compounds (except 2g) were suitable in the reaction. As for the aromatic aldehydes, electron-donating group on the phenyl ring had a better reactivity than those with electron-withdrawing group. When 1a was treated with the mixture of 1.5 equiv of 4nitrobenzaldehyd (2c) and 1.5 equiv of 4-methoxybenzaldehyde (2d) in the presence of 5 equiv of BF3:Et2O, the molar ratio of product 3ac:3ad was about 1:3 (see Supporting Information). Heteroatomic aldehydes were also tested (Table 2, entries 5-7). Although 2-thenaldehyde 2e gave excellent yield of 3ae (93%), 2-furaldehyde 2f only gave moderate yield of 3af (56%) due to the generation of some byproducts, and nicotinaldehyde **2g** failed in this reaction.  $\alpha,\beta$ -Unsaturated aldehydes and aliphatic aldehydes could also be used as the dipolarophiles to afford the corresponding products in high yields (Table 2, entries 8-12). The ketonic compounds are also applicable in this transformation with 15 equiv of BF<sub>3</sub>·Et<sub>2</sub>O (Table 2, entries 13 and 14).

Table 2. Substrate Scope for the BF<sub>3</sub>·Et<sub>2</sub>O-Catalyzed Formal [3 + 2] Reaction<sup>a</sup>

| entry           | carbonyls                         | product | time<br>(min) | yield (%) <sup>b</sup> |  |
|-----------------|-----------------------------------|---------|---------------|------------------------|--|
| 1               | 2а СНО                            | 3aa     | 25            | 88                     |  |
| 2               | <b>2b</b><br>сі—Сно               | 3ab     | 20            | 92                     |  |
| 3               | <b>2c</b><br>O <sub>2</sub> N—CHO | 3ac     | 30            | 74                     |  |
| 4               | 2d<br>MeO—CHO                     | 3ad     | 30            | 90                     |  |
| 5               | 2e S CHO                          | 3ae     | 35            | 93                     |  |
| 6               | 2f CHO                            | 3af     | 60            | 56                     |  |
| 7               | <b>2g</b> N= CHO                  | 3ag     | 60            | 0                      |  |
| 8               | 2h CHO                            | 3ah     | 25            | 86                     |  |
| $9^c$           | 2i CHO                            | 3ai     | 55            | 94                     |  |
| $10^c$          | 2ј 🗡 сно                          | 3aj     | 15            | 90                     |  |
| 11 <sup>c</sup> | <b>2k</b> —сно                    | 3ak     | 15            | 92                     |  |
| 12 <sup>d</sup> | <b>2</b> 1 H                      | 3al     | 60            | 70                     |  |
| 13 <sup>e</sup> | 2m 🗎                              | 3am     | 35            | 83                     |  |
| 14 <sup>f</sup> | <b>2</b> n =0                     | 3an     | 25            | 76                     |  |

<sup>a</sup>Unless notification, the reactions were carried out with the molar ratio of **1a** (17.8 mg)/2/BF<sub>3</sub>·Et<sub>2</sub>O = 1:1.5:5 in 2.5 mL of dry chlorobenzene at room temperature. <sup>b</sup>Isolated yield. <sup>c</sup>**1a**/2/BF<sub>3</sub>·Et<sub>2</sub>O = 1:3:5. <sup>d</sup>**1a**/paraformaldehyde/BF<sub>3</sub>·Et<sub>2</sub>O = 1:5:5. <sup>e</sup>**1a**/2m/BF<sub>3</sub>·Et<sub>2</sub>O = 1:4:15. <sup>f</sup>**1a**/2n/BF<sub>3</sub>·Et<sub>2</sub>O = 1:2:15.

We next turned to study the applicability of other aziridinofullerenes in this conversion by performing the reaction of them with 4-chlorobenzaldehyde (Table 3). The aziridinofullerene with an electron-withdrawing group on the phenyl ring had a better reactivity than that with an electron-donating group. *N*-Methylaziridinofullerene 1d also worked well to give the corresponding product 3db in 90% yield.

Table 3. Study of the Reaction Scope by Variation of Aziridinofullerene $^a$ 

| entry | substrate | R                 | product | time  | yield    |
|-------|-----------|-------------------|---------|-------|----------|
|       |           |                   |         | (min) | $(\%)^b$ |
| 1     | 1b        | MeO-              | 3bb     | 100   | 82       |
| 2     | 1c        | O <sub>2</sub> N- | 3cb     | 25    | 94       |
| 3     | 1d        | Me —              | 3db     | 25    | 90       |

<sup>a</sup>The reactions were carried out with the molar ratio of 1 (0.02 mmol):2b:BF<sub>3</sub>·Et<sub>2</sub>O = 1:1.5:5 in 2.5 mL of dry chlorobenzene at room temperature. <sup>b</sup>Isolated yield.

Among the above investigated alkyl aldehydes, the normal [3 + 2] products were obtained as expected. When the propionaldehyde **2o**, which possessed two  $\alpha$ -H atoms, was subjected to this reaction, TLC indicated that full conversion to a single product had occurred. However, the NMR analysis showed the **3ao** was generated as the main product along with a small amount of byproduct **4** (**3ao**/**4** = 89/11) (Scheme 2).

Scheme 2. Reaction of N-Tosylaziridinofullerene 1a with Propionaldehyde 2o

The generation of 4 could be explained by the cycloaddition reaction of 1a with (E)-2-methylpent-2-enal, which could be generated from the self-condensation of 20 under BF3·Et2O conditions. In order to suppress the side reaction, the amount of 20 was raised to 15 equiv. Contrary to our expectations, the reaction proceeded much more slowly than 3 equiv of 20 and the content of 4 increased greatly (3ao/4 = 57/43) (Scheme 2). In order to verify whether the product 4 was formed through the reaction of **3ao** with (E)-2-methylpent-2-enal, that is, whether the BF<sub>3</sub>·Et<sub>2</sub>O-catlyzed formal [3 + 2] reaction of 1 with carbonyls was reversible, the reaction of 3aj with 2d in the presence of 5 equiv of BF<sub>3</sub>·Et<sub>2</sub>O was carried out. <sup>16</sup> No reaction occurred and 3aj was totally recovered, which indicated that such BF<sub>3</sub>·Et<sub>2</sub>O-catlyzed formal [3 + 2] reaction of aziridinofullerene with carbonyls was irreversible (see Supporting Information). This also demonstrated that 1a reacted with 2o and (E)-2-methylpent-2-enal to provide **3ao** and **4**, respectively.

To further identify the structure of 4, the reaction of 1a with 1.5 equiv of (*E*)-2-methylpent-2-enal in the presence of 5 equiv of  $BF_3 \cdot Et_2O$  was examined. Product 4 was obtained as the sole product in 95% yield.

Although the [3 + 2] reaction of aziridines with carbonyl compounds has been well documented,  $^{3d,15,17}$  the reactivity of those compounds with a carbonyl group such as DMF, Nmethylpyrrolidin-2-one, ethyl formate, and dimethyl carbonate had never been investigated in this kind of transformation. N-Methylpyrrolidin-2-one, DMF, and dimethyl carbonate showed no reactivity under standard conditions. However, when ethyl formate 5 was employed in the reaction with the molar ratio of 1a:5:BF<sub>3</sub>·Et<sub>2</sub>O as 1:10:20, an unprecedented fullerooxazole product 6 without substituent at the 2-position was isolated in 78% yield. Although several methods have been reported to prepare the fullerooxazole derivatives, <sup>13e,18</sup> all of them failed to give the simplest fullerooxazole 6. The oxazole skeleton bearing a hydrogen atom at 2-position has been reported to undergo various transformations to generate new C-X (X = C, N, O, or S) bonds though C-H functionalization. 19 It could be predicted that fullerooxazole 6 could also be used as a diverse platform for further functionalization to prepare many fullerene derivatives which were difficult to synthesize from C<sub>60</sub> directly.

Scheme 3. Reaction of N-Tosylaziridinofullerene 1a with Ethyl Formate 5

The structures of  $C_{60}$ -fused oxazolidines 3 were fully assigned on the basis of their MALDI-TOFMS,  $^1\mathrm{H}$  NMR,  $^{13}\mathrm{C}$  NMR, and UV–vis spectra. With 3ab taken as an example, the TOFMS spectrum of 3ab showed the [M+Na]<sup>+</sup> peak at m/z 1052.0118. The  $^1\mathrm{H}$  NMR spectrum of 3ab displayed a distinct singlet at 7.62 ppm for methine hydrogen. The  $^{13}\mathrm{C}$  NMR spectrum of 3ab exhibited 56 signals (51 × 1C, 4 × 2C, and 1 × 3C) for the sp<sup>2</sup>-C of the  $C_{60}$  skeleton and aromatic-C in the range of 135.68–150.17 ppm, four signals (each 2C) for the aryl-C of phenyl ring in the range of 128.09–130.01 ppm, two peaks at 79.22 and 98.81 ppm for the sp<sup>3</sup>-C of the  $C_{60}$  cage, and a characteristic peak at 92.26 ppm for the NCH(R)O moiety, agreeing with  $C_1$  symmetry of its molecular structure.

In summary, we have developed an efficient BF $_3$ ·Et $_2$ O-catalyzed formal [3 + 2] reaction of aziridinofullerenes with a variety of carbonyl compounds for the preparation of novel C $_{60}$ -fused oxazolidines. Moreover, ethyl formate could be used as a dipolarophile in the reaction to generate the simplest fullerooxazole without substituent at the 2-position.

# **EXPERIMENTAL SECTION**

General Procedure for the BF<sub>3</sub>·Et<sub>2</sub>O-Catalyzed Formal [3 + 2] Cycloaddition Reaction of Aziridinofullerenes with Carbonyl Compounds. Aziridinofullerenes 1 (0.02 mmol) and carbonyl coumpounds 2 (2a–h: 0.03 mmol; 2i–k and (E)-2-methylpent-2-enal: 0.06 mmol; 2l: 0.1 mmol; 2m: 0.08 mmol; 2n: 0.04 mmol) were dissolved in 2.5 mL of dry chlorobenzene. Then, BF<sub>3</sub>·Et<sub>2</sub>O (for 2a–l, 0.1 mmol, 12  $\mu$ L; for 2m and 2n, 0.3 mmol, 37  $\mu$ L) was added to the solution, and the mixture was stirred at room temperature until the disappearance of 1 determined by TLC. The solvent was removed in vacuo, and the residue was purified on a silica gel column using CS<sub>2</sub>/toluene as the eluent to give the products 3.

3aa (brown solid, 17.6 mg, 88%, mp >300 °C):  $^{1}\mathrm{H}$  NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.13–8.15 (m, 2H), 7.95 (d, J = 8.3 Hz, 2H), 7.68 (s, 1H), 7.44–7.50 (m, 3H), 7.34 (d, J = 8.1 Hz, 2H), 2.48 (s, 3H);  $^{13}\mathrm{C}$  NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.36, 148.36, 148.14, 147.99, 147.92, 146.62, 146.57, 146.50, 146.34, 146.19, 146.15, 146.09, 146.01, 145.80, 145.31, 145.28, 145.16, 145.03, 144.96, 144.89, 144.79, 144.58, 144.52, 144.43, 144.37, 144.29, 144.04, 142.80, 142.76, 142.71, 142.68, 142.65, 142.51, 142.50, 142.32, 142.29, 142.25, 142.04, 141.98, 141.87, 141.40, 141.11, 141.08, 139.85, 139.51, 139.43, 138.53, 138.11, 138.09, 137.42, 137.39, 137.01, 136.71, 129.74, 129.71, 128.57, 128.31, 128.07, 98.68, 92.88, 79.17, 21.82; UV–vis (CHCl<sub>3</sub>)  $\lambda_{\max}$ /nm 257, 318, 453, 684; HRMS (MALDI-TOFMS) m/z [M+Na]+ calcd for  $\mathrm{C_{74}H_{13}NNaO_{3}S}$  1018.0514, found 1018.0513.

3ab (brown solid, 18.9 mg, 92%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>) δ 8.08 (d, J = 8.3 Hz, 2H), 7.94 (d, J = 8.4 Hz, 2H), 7.62 (s, 1H), 7.45 (d, J = 8.5 Hz, 2H), 7.35 (d, J = 8.1 Hz, 2H), 2.49 (s, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>) δ 150.17, 148.23, 148.08, 147.70, 146.64, 146.57, 146.56, 146.46, 146.44, 146.28, 146.24, 146.18, 146.10, 145.92, 145.90, 145.37, 145.25, 145.12, 145.05, 144.88, 144.83, 144.63, 144.56, 144.51, 144.43, 144.23, 144.02, 142.89, 142.87, 142.84, 142.81, 142.75, 142.72, 142.58, 142.54, 142.39, 142.35, 142.30, 142.10, 142.07, 141.96, 141.42, 141.26, 141.23, 141.12, 139.93, 139.65, 139.55, 138.32, 138.20, 138.14, 137.45, 137.41, 136.59, 136.06, 135.68, 130.01, 129.82, 128.63, 128.09, 98.81, 92.26, 79.22, 21.85; UV–vis (CHCl<sub>3</sub>)  $λ_{\text{max}}/\text{nm}$  257, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for  $C_{74}\text{H}_{12}\text{CINNaO}_{3}\text{S}$  1052.0124, found 1052.0118.

3ac (brown solid, 15.4 mg, 74%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.40 (d, J = 9.0 Hz, 2H), 8.36 (d, J = 8.9 Hz, 2H), 7.97 (d, J = 8.3 Hz, 2H), 7.71 (s, 1H), 7.38 (d, J = 8.2 Hz, 2H), 2.50 (s, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.01, 148.95, 148.25, 148.09, 147.60, 147.29, 146.65, 146.59, 146.48, 146.45, 146.38, 146.31, 146.30, 146.25, 146.21, 146.12, 145.98, 145.92, 145.36, 145.27, 145.22, 145.13, 145.05, 144.96, 144.93, 144.79, 144.60, 144.49, 144.39, 144.35, 143.84, 143.63, 142.93, 142.88, 142.85, 142.76, 142.73, 142.55, 142.51, 142.37, 142.33, 142.27, 142.11, 142.08, 141.97, 141.37, 141.18, 141.08, 141.06, 139.94, 139.70, 139.69, 138.25, 138.15, 138.08, 137.41, 137.15, 136.14, 129.93, 129.56, 128.10, 123.58, 99.09, 91.85, 79.10, 21.87; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}$ /nm 257, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for  $C_{74}H_{12}N_2NaO_5$ S 1063.0365, found 1063.0371.

3ad (brown solid, 18.4 mg, 90%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.99 (d, J = 8.4 Hz, 2H), 7.90 (d, J = 8.4 Hz, 2H), 7.58 (s, 1H), 7.32 (d, J = 8.3 Hz, 2H), 6.94 (d, J = 8.6 Hz, 2H), 3.86 (s, 3H), 2.48 (s, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  160.75, 150.29, 148.44, 148.11, 148.00, 147.97, 146.65, 146.54, 146.48, 146.33, 146.16, 146.14, 146.06, 145.99, 145.79, 145.77, 145.34, 145.25, 145.13, 145.05, 145.01, 144.94, 144.77, 144.66, 144.57, 144.52, 144.38, 144.10, 142.77, 142.74, 142.69, 142.66, 142.63, 142.51, 142.48, 142.30, 142.27, 142.23, 142.00, 141.98, 141.86, 141.38, 141.22, 141.18, 141.08, 139.84, 139.53, 139.33, 138.55, 138.07, 138.01, 137.44, 137.38, 136.77, 129.95, 129.61, 128.72, 128.02, 113.64, 98.47, 92.68, 79.19, 55.18, 21.79; UV–vis (CHCl<sub>3</sub>)  $\lambda_{\text{max}}$ /nm 257, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]+ calcd for  $C_{75}H_{15}$ NNaO<sub>4</sub>S 1048.0619, found 1048.0612.

3ae (brown solid, 18.6 mg, 93%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.95 (d, J = 8.3 Hz, 2H), 7.81 (s, 1H), 7.77 (d, J = 3.4 Hz, 1H), 7.46 (d, J = 4.9 Hz, 1H), 7.34 (d, J = 8.2 Hz, 2H), 7.11 (dd, J = 4.9, 3.4 Hz, 1H), 2.49 (s, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.39, 148.27, 148.08, 147.94, 147.75, 146.51, 146.49, 146.45, 146.32, 146.28, 146.17, 146.14, 146.07, 146.03, 145.96, 145.82, 145.73, 145.33, 145.23, 145.09, 144.98, 144.91, 144.89, 144.78, 144.71, 144.53, 144.50, 144.41, 144.32, 144.25, 144.23, 144.05, 142.74, 142.72, 142.67, 142.65, 142.62, 142.44, 142.32, 142.27, 142.21, 142.01, 141.89, 141.78, 141.51, 141.33, 141.16, 141.12, 141.03, 139.78, 139.51, 139.33, 138.33, 138.06, 137.99, 137.36, 137.13, 136.78, 129.72, 129.43, 128.12, 128.02, 126.85, 99.07, 90.17, 78.90, 21.82; UV—vis (CHCl<sub>3</sub>)  $\lambda$ max/nm 256, 319, 454, 684; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for  $C_{72}H_{11}$ NNaO<sub>3</sub>S<sub>2</sub> 1024.0078, found 1024.0111.

3af (brown solid, 11.0 mg, 56%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>-CS<sub>2</sub>)  $\delta$  7.91 (d, J = 8.3 Hz, 2H), 7.57 (br, 1H), 7.55 (s,

1H), 7.33 (d, J=8.3 Hz, 2H), 7.12 (d, J=3.4 Hz, 1H), 6.55 (dd, J=3.3, 1.9 Hz, 1H), 2.47 (s, 3H);  $^{13}{\rm C}$  NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.06, 150.05, 148.19, 148.06, 147.94, 147.84, 146.58, 146.55, 146.47, 146.40, 146.26, 146.19, 146.18, 146.14, 146.09, 146.04, 145.85, 145.43, 145.28, 145.17, 145.14, 145.03, 144.88, 144.83, 144.75, 144.65, 144.61, 144.54, 144.49, 144.38, 144.36, 143.98, 143.15, 142.85, 142.84, 142.80, 142.72, 142.71, 142.54, 142.50, 142.45, 142.36, 142.33, 142.09, 142.05, 141.91, 141.45, 141.38, 141.24, 139.87, 139.67, 139.30, 138.23, 138.00, 137.92, 137.88, 137.20, 129.77, 128.12, 112.65, 110.95, 99.27, 87.09, 78.80, 21.82; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  256, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]+ calcd for C<sub>72</sub>H<sub>11</sub>NNaO<sub>4</sub>S 1008.0306, found 1008.0294.

3ah (brown solid, 17.5 mg, 86%, mp >300 °C): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>-CS<sub>2</sub>)  $\delta$  7.91 (d, J = 8.3 Hz, 2H), 7.46 (d, J = 7.2 Hz, 2H), 7.37 (d, J = 15.8 Hz, 1H), 7.35 (t, J = 7.4 Hz, 2H), 7.30 (t, J = 7.2Hz, 1H), 7.29 (d, I = 8.3 Hz, 2H), 7.02 (dd, I = 5.2, 1.2 Hz, 1H), 6.85 (dd, J = 15.8, 5.2 Hz, 2H), 2.45 (s, 3H); <sup>13</sup>C NMR (125 MHz,  $CDCl_3-CS_2$ )  $\delta$  150.22, 148.70, 148.09, 147.94, 147.86, 146.50, 146.45, 146.39, 146.32, 146.17, 146.11, 146.10, 146.05, 145.98, 145.86, 145.33, 145.31, 145.19, 145.17, 145.09, 145.03, 145.02, 144.96, 144.71, 144.55, 144.50, 144.40, 144.08, 142.79, 142.74, 142.72, 142.62, 142.60, 142.47, 142.45, 142.30, 142.26, 142.22, 142.07, 141.96, 141.87, 141.49, 141.39, 141.36, 141.30, 139.83, 139.70, 139.19, 138.31, 138.11, 137.70, 137.62, 137.32, 136.51, 136.39, 135.47, 129.57, 128.79, 128.76, 128.19, 127.34, 125.02, 98.65, 92.48, 78.84, 21.75; UV–vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  256, 318, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na] calcd for C<sub>76</sub>H<sub>15</sub>NNaO<sub>3</sub>S 1044.0670, found 1044.0682.

3ai (brown solid, 18.0 mg, 94%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.90 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.1 Hz, 2H), 6.82 (d, J = 5.4 Hz, 1H), 6.52–6.61 (m, 1H), 6.30–6.35 (m, 1H), 2.47 (s, 3H), 1.97 (d, J = 6.6 Hz, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.45, 148.93, 148.11, 147.97, 147.95, 146.50, 146.46, 146.42, 146.35, 146.19, 146.14, 146.08, 146.07, 146.02, 145.91, 145.36, 145.34, 145.18, 145.16, 145.10, 145.05, 145.02, 144.98, 144.75, 144.56, 144.55, 144.46, 144.42, 144.40, 144.10, 142.79, 142.75, 142.73, 142.68, 142.63, 142.47, 142.45, 142.30, 142.27, 142.23, 142.08, 141.97, 141.87, 141.49, 141.40, 141.37, 141.30, 139.83, 139.68, 139.20, 138.37, 138.10, 137.68, 137.62, 137.27, 136.53, 133.67, 129.54, 128.12, 127.64, 98.55, 92.69, 78.68, 21.78, 17.80; UV–vis (CHCl<sub>3</sub>)  $\lambda$ <sub>max</sub>/nm 256, 318, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for C<sub>71</sub>H<sub>13</sub>NNaO<sub>3</sub>S 982.0514, found 982.0501.

3aj (brown solid, 17.6 mg, 90%, mp >300 °C):  $^{1}\mathrm{H}$  NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.87 (d, J = 8.3 Hz, 2H), 7.27 (d, J = 8.3 Hz, 2H), 6.54 (s, 1H), 2.44 (s, 3H), 1.66 (s, 9H);  $^{13}\mathrm{C}$  NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  151.05, 149.22, 148.57, 148.10, 147.98, 146.44, 146.39, 146.37, 146.09, 146.06, 146.02, 145.94, 145.93, 145.72, 145.65, 145.34, 145.15, 145.08, 145.02, 144.99, 144.88, 144.69, 144.65, 144.57, 144.52, 144.36, 144.16, 144.12, 143.90, 143.88, 142.80, 142.78, 142.76, 142.68, 142.62, 142.60, 142.51, 142.39, 142.29, 142.21, 142.01, 142.08, 141.74, 141.36, 141.15, 140.91, 140.47, 139.69, 139.50, 139.31, 138.55, 137.89, 137.24, 137.09, 136.54, 129.75, 127.61, 102.68, 99.28, 80.25, 38.88, 28.06, 21.77; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/\rm nm$  257, 319, 454, 684; HRMS (MALDI-TOFMS) m/z [M+Na] + calcd for C<sub>72</sub>H<sub>17</sub>NNaO<sub>3</sub>S 998.0827, found 998.0834.

3ak (brown solid, 17.7 mg, 92%, mp >300 °C):  $^{1}{\rm H}$  NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.94 (d, J = 8.2 Hz, 2H), 7.33 (d, J = 8.2 Hz, 2H), 6.31 (d, J = 9.5 Hz, 2H), 3.53–3.62 (m, 1H), 2.46 (s, 3H), 1.57 (d, J = 6.8 Hz, 3H), 1.52 (d, J = 6.6 Hz, 3H);  $^{13}{\rm C}$  NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  151.93, 149.54, 148.19, 148.05, 146.56, 146.48, 146.41, 146.29, 146.21, 146.19, 146.18, 146.16, 146.08, 146.03, 145.40, 145.30, 145.19, 145.10, 145.01, 144.83, 144.62, 144.57, 144.40, 144.33, 144.24, 143.67, 142.87, 142.80, 142.77, 142.73, 142.69, 142.52, 142.48, 142.27, 142.25, 142.10, 142.05, 141.96, 141.41, 141.35, 141.31, 140.64, 139.90, 139.75, 139.64, 139.14, 138.09, 137.86, 137.38, 137.29, 136.21, 129.72, 128.02, 100.22, 98.43, 79.13, 33.63, 21.81, 19.61, 18.97; UV–vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  257, 319, 452, 683; HRMS (MALDI-TOFMS) m/z [M+Na] $^+$  calcd for C<sub>71</sub>H<sub>15</sub>NNaO<sub>3</sub>S 984.0670, found 984.0653.

**3al** (brown solid, 12.9 mg, 70%, mp >300 °C): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.99 (d, J = 8.3 Hz, 2H), 7.37 (d, J = 8.3 Hz, 2H), 6.25 (s, 2H), 2.48 (s, 3H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$ 

148.26, 148.09, 146.99, 146.88, 146.60, 146.53, 146.28, 146.24, 146.21, 146.09, 145.52, 145.26, 145.23, 144.68, 144.63, 144.58, 144.57, 142.88, 142.86, 142.75, 142.52, 142.33, 142.30, 142.08, 141.47, 141.44, 139.89, 139.12, 138.02, 137.50, 137.39, 129.75, 128.47, 98.51, 80.12, 78.74, 21.83; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  257, 319, 453, 684; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for C<sub>68</sub>H<sub>9</sub>NNaO<sub>3</sub>S 942.0201, found 942.0198.

3am (brown solid, 15.7 mg, 83%, mp >300 °C):  $^{1}{\rm H}$  NMR (500 MHz, CDCl3-CS2)  $\delta$  7.89 (d, J = 8.3 Hz, 2H), 7.28 (d, J = 8.3 Hz, 2H), 2.45 (s, 3H), 2.44 (s, 6H);  $^{13}{\rm C}$  NMR (125 MHz, CDCl3-CS2) (all 2C unless indicated)  $\delta$  149.63, 148.96, 148.01, 147.92, 146.42, 146.39, 146.15, 145.98, 145.96, 145.21, 144.99, 144.97, 144.56, 144.53, 144.34, 143.39, 142.73, 142.69, 142.58, 142.46, 142.20, 142.18, 141.75, 141.63, 141.37, 139.79, 137.94, 136.94, 136.69, 129.35, 127.79, 99.38, 96.87, 80.06, 28.47, 21.66; UV-vis (CHCl3)  $\lambda_{\rm max}/{\rm nm}$  257, 319, 452, 684; HRMS (MALDI-TOFMS) m/z [M+Na]+ calcd for  ${\rm C_{70}H_{13}NNaO_{3}S}$  970.0514, found 970.0501.

3an (brown solid, 15.1 mg, 76%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.92 (d, J = 8.3 Hz, 2H), 7.29 (d, J = 8.3 Hz, 2H), 2.75–2.93 (m, 4H), 2.45 (s, 3H), 1.87–2.17 (m, 5H), 1.51–1.64 (m, 1H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  149.81, 149.24, 148.21, 148.12, 146.66, 146.62, 146.56, 146.34, 146.15, 146.14, 145.40, 145.19, 145.16, 144.74, 144.72, 144.65, 143.63, 142.90, 142.88, 142.75, 142.65, 142.38, 142.34, 141.91, 141.79, 141.52, 140.29, 139.96, 138.10, 137.15, 136.97, 129.56, 127.77, 101.76, 96.94, 80.54, 36.89, 25.17, 24.37, 21.72; UV–vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  257, 319, 452, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for C<sub>73</sub>H<sub>17</sub>NNaO<sub>3</sub>S 1010.0827, found 1010.0834.

4: (brown solid, 18.8 mg, 95%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.90 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.2 Hz, 2H), 6.70 (s, 1H), 6.49 (t, J = 7.2 Hz, 1H), 2.48 (s, 3H), 2.27–2.36 (m, 2H), 2.04 (s, 3H), 1.18 (t, J = 7.6 Hz, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.31, 148.55, 148.23, 148.12, 147.99, 146.58, 146.52, 146.48, 146.40, 146.22, 146.16, 146.12, 146.06, 146.03, 145.97, 145.47, 145.44, 145.22, 145.14, 145.07, 145.04, 144.98, 144.92, 144.79, 144.62, 144.56, 144.47, 144.42, 144.17, 142.79, 142.74, 142.71, 142.64, 142.52, 142.51, 142.31, 142.29, 142.25, 142.10, 142.02, 141.86, 141.44, 141.33, 141.31, 141.22, 139.88, 139.69, 139.18, 138.01, 137.90, 137.84, 137.22, 135.18, 130.00, 129.50, 128.35, 97.94, 95.63, 79.29, 29.96, 21.84, 13.55, 12.35; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}$ /nm 256, 318, 453, 684; HRMS (MALDITOFMS) m/z [M+Na]+ calcd for C<sub>73</sub>H<sub>17</sub>NNaO<sub>3</sub>S 1010.0827, found 1010.0828.

**3bb** (brown solid, 17.1 mg, 82%, mp >300 °C):  $^{1}$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.04 (d, J = 8.3 Hz, 2H), 7.91 (d, J = 8.8 Hz, 2H), 7.54 (s, 1H), 7.42 (d, J = 8.3 Hz, 2H), 6.97 (d, J = 8.8 Hz, 2H), 3.89 (s, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  163.39, 150.08, 148.05, 147.96, 147.90, 147.61, 146.48, 146.44, 146.42, 146.29, 146.28, 146.11, 146.08, 146.02, 145.93, 145.78, 145.75, 145.19, 145.09, 144.96, 144.93, 144.88, 144.85, 144.68, 144.49, 144.41, 144.38, 144.30, 143.92, 142.74, 142.72, 142.69, 142.65, 142.61, 142.58, 142.44, 142.40, 142.23, 142.21, 142.15, 141.95, 141.92, 141.79, 141.29, 141.15, 141.10, 141.08, 139.78, 139.50, 139.35, 138.01, 137.86, 137.29, 137.26, 136.39, 135.95, 135.49, 132.66, 130.06, 129.88, 128.48, 114.14, 98.55, 92.05, 79.01, 55.48; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\text{max}}$ /nm 257, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for C<sub>74</sub>H<sub>12</sub>ClNNaO<sub>4</sub>S 1068.0073, found 1068.0062.

3cb (brown solid, 19.9 mg, 94%, mp >300 °C): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.36 (d, J = 8.9 Hz, 2H), 8.19 (d, J = 8.8 Hz, 2H), 8.03 (d, J = 8.4 Hz, 2H), 7.56 (s, 1H), 7.43 (d, J = 8.4 Hz, 2H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  150.30, 148.92, 148.19, 148.09, 147.55, 147.36, 146.79, 146.65, 146.64, 146.47, 146.46, 146.41, 146.30, 146.28, 146.26, 146.18, 146.13, 145.98, 145.96, 145.36, 145.22, 145.20, 145.10, 144.92, 144.71, 144.60, 144.57, 144.29, 144.21, 143.82, 143.54, 142.94, 142.90, 142.87, 142.84, 142.74, 142.49, 142.46, 142.41, 142.33, 142.31, 142.10, 142.08, 141.91, 141.34, 141.20, 141.12, 140.85, 140.10, 139.73, 139.46, 138.30, 138.02, 137.27, 136.87, 136.81, 136.57, 134.59, 130.03, 128.95, 128.73, 124.21, 98.80, 92.02, 79.17; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\text{max}}/\text{nm}$  257, 319, 452, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for  $C_{73}\text{H}_{9}\text{ClN}_{3}\text{NaO}_{9}$ S 1082.9818, found 1082.9811.

3db (brown solid, 17.1 mg, 90%, mp >300 °C):  $^{1}{\rm H}$  NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.10 (d, J = 8.3 Hz, 2H), 7.47 (d, J = 8.4 Hz, 2H), 7.37 (s, 1H), 3.56 (s, 3H);  $^{13}{\rm C}$  NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  149.97, 148.17, 148.02, 147.81, 146.71, 146.61, 146.58, 146.42, 146.36, 146.35, 146.23, 146.22, 146.14, 146.08, 145.83, 145.75, 145.37, 145.23, 145.16, 145.09, 145.06, 144.99, 144.95, 144.76, 144.52, 144.48, 144.35, 144.23, 143.84, 142.88, 142.85, 142.75, 142.65, 142.51, 142.42, 142.27, 142.24, 142.11, 141.96, 141.85, 141.62, 141.60, 141.09, 141.05, 140.05, 139.61, 139.60, 138.43, 137.33, 137.27, 136.94, 136.37, 136.22, 135.30, 129.82, 128.65, 98.50, 91.74, 79.67, 40.96; UV—vis (CHCl<sub>3</sub>)  $\lambda_{\rm max}/{\rm nm}$  257, 319, 453, 683; HRMS (MALDI-TOFMS) m/z M $^+$  calcd for  $C_{68}H_8{\rm CINO}_3{\rm S}$  952.9913, found 952.9933.

BF<sub>3</sub>·Et<sub>2</sub>O-Catalyzed Reaction of Aziridinofullerene 1a with Propionaldehyde 2o. Aziridinofullerene 1a (17.8 mg, 0.02 mmol) and propionaldehyde 2o (0.06 or 0.3 mmol) were dissolved in 2.5 mL of dry chlorobenzene. Then, BF<sub>3</sub>·Et<sub>2</sub>O (0.1 mmol, 12  $\mu$ L) was added to the solution, and the mixture was stirred at room temperatrue until the disappearance of 1a determined by TLC. The solvent was evaporated *in vacuo*, and the residue was purified on a silica gel column using CS<sub>2</sub>/toluene as the eluent to give the inseparable products 3ao and 4 (when 1a/2o/BF<sub>3</sub>·Et<sub>2</sub>O = 1:3:5, 16.8 mg, 88%, 3ao/4 = 89:11; when 1a/2o/BF<sub>3</sub>·Et<sub>2</sub>O = 1:15:5, 15.6 mg, 81%, 3ao/4 = 57:43).

3ao (brown solid, mp >300 °C):  $^1$ H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  7.93 (d, J = 8.3 Hz, 2H), 7.33 (d, J = 8.2 Hz, 2H), 6.49 (dd, J = 8.7, 4.6 Hz, 1H), 3.01–3.10 (m, 1H), 2.78–2.87 (m, 1H), 2.47 (s, 3H), 1.49 (t, J = 7.4 Hz, 3H);  $^{13}$ C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  151.60, 149.53, 148.23, 148.12, 148.08, 146.59, 146.53, 146.44, 146.42, 146.28, 146.23, 146.18, 146.16, 146.12, 146.04, 145.46, 145.29, 145.21, 145.13, 145.10, 145.06, 144.96, 144.87, 144.64, 144.61, 144.51, 144.50, 144.45, 144.43, 144.25, 142.88, 142.83, 142.80, 142.70, 142.54, 142.51, 142.36, 142.33, 142.29, 142.14, 142.07, 141.96, 141.59, 141.48, 141.20, 139.84, 139.74, 139.61, 138.70, 138.04, 137.91, 137.34, 137.21, 136.13, 129.76, 127.85, 98.88, 95.80, 78.63, 30.35, 21.80, 10.17; UV–vis (CHCl<sub>3</sub>)  $\lambda$ <sub>max</sub>/nm 256, 319, 453, 683; HRMS (MALDI-TOFMS) m/z [M+Na]<sup>+</sup> calcd for C<sub>70</sub>H<sub>13</sub>NNaO<sub>3</sub>S 970.0514, found 970.0524.

BF<sub>3</sub>·Et<sub>2</sub>O-Catalyzed Reaction of Aziridinofullerene 1a with Ethyl Formate 5. Aziridinofullerene 1a (17.8 mg, 0.02 mmol) and ethyl formate 5 (14.8 mg, 0.2 mmol) were dissolved in 2.5 mL of dry chlorobenzene. Then, BF<sub>3</sub>·Et<sub>2</sub>O (0.4 mmol, 49  $\mu$ L) was added to the solution, and the mixture was stirred at room temperature until the disappearance of 1a determined by TLC. The solvent was evaporated *in vacuo*, and the residue was purified on a silica gel column using CS<sub>2</sub>/ toluene as the eluent to give the products 6.

6 (brown solid, 11.9 mg, 78%, mp >300 °C): <sup>1</sup>H NMR (500 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>)  $\delta$  8.03 (s, 1H); <sup>13</sup>C NMR (125 MHz, CDCl<sub>3</sub>–CS<sub>2</sub>) (all 2C unless indicated)  $\delta$  156.50, 148.27, 147.86, 147.16, 146.48, 146.36, 146.18, 146.13, 145.90, 145.57, 145.48, 145.31, 145.21, 144.82, 144.88, 144.29, 142.89, 142.83, 142.78, 142.43, 142.34, 142.26, 142.18, 142.05, 141.96, 140.52, 139.79, 137.72, 136.22, 96.55, 90.96; UV–vis (CHCl<sub>3</sub>)  $\lambda$ <sub>max</sub>/nm 256, 317, 454, 611, 684; HRMS (MALDI-TOFMS) m/z M<sup>+</sup> calcd for C<sub>61</sub>HNO 763.0058, found 763.0049.

## ASSOCIATED CONTENT

### S Supporting Information

Control experiments, UV—vis spectrum of **3db**, and <sup>1</sup>H and <sup>13</sup>C NMR spectra of the products. This material is available free of charge via the Internet at http://pubs.acs.org.

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#### Notes

The authors declare no competing financial interest.

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- conducted the reaction of **3ao** (a mixture with **4**, **3ao**/**4** = 89/11) with 3 equiv of (E)-2-methylpent-2-enal in the presence of 5 equiv of BF<sub>3</sub> × Et<sub>2</sub>O. After stirring for 5 h at room temperature no change of the molar ratio of **3ao**/**4** was observed. This result proved the irreversibility of the reaction directly.
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